

Investigation on the Seroprevalence of *Toxoplasmosis*, *Listeriosis* and *Brucellosis* in Goats living in the region of Van, Turkey

Mehmet KARACA¹ Cahit BABÜR² Bekir ÇELEBİ² Hasan Altan AKKAN¹,
Mehmet TÛTÛNCÛ³ İhsan KELEŞ¹ Barış Atalay USLU⁴ Selçuk KILIÇ²

¹Dept. of Internal Medicine, Faculty of Veterinary, University of Yuzuncu Yıl, Van-Turkey

²Refik Saydam National Institute of Hygiene, Dept. of Communicable Diseases Research, Ankara-Turkey

³Van Health Services Vocational Collage, University of Yuzuncu Yıl, Van-Turkey

⁴Dept. of Reproduction on Artificial Insemination, Faculty of Veterinary, University of Yuzuncu Yıl, Van-Turkey

Sorumlu Araştırmacı, 0505 3562949 vetmkaraca@hotmail.com

Summary: This study was carried out on goat sera to determine the seroprevalence of listeriosis, toxoplasmosis and brucellosis in the province of Van. Sabin Feldman Dye Test (SFDT), Osebold Agglutination Test (OAT) and Micro Agglutination Test (MAT) were applied to determine antibodies developed against *Toxoplasma gondii*, *Listeria monocytogenes* and *Brucella melitensis* respectively. Out of the 98 sera, 80.61% toxoplasmosis, 34.69% listeriosis and 6.12% brucellosis were evaluated as positive. This results indicate that *Toxoplasmosis* is much more widespread than *Listeriosis*, and *Brucellosis* in goats living in the province of Van.

Key words: *Toxoplasma*, *Listeria*, *Brucella*, serology, seroprevalence, goats

Van Yöresi Keçilerinde *Toxoplasmozis*, *Listeriozis* ve *Brucellozis*'in Seroprevalansı Üzerine Araştırmalar

Özet: Bu çalışma Van yöresi keçilerinde *toxoplasmosis*, *listeriosis* ve *brucellosis* seroprevalansını belirlemek için yapıldı. *Toxoplasma gondii*, *Listeria monocytogenes* ve *Brucella melitensis*'e karşı oluşan antikorları belirlemek için sırası ile Sabin Feldman Dye Testi (SFDT), Osebold Agglutination Testi (OAT) ve Micro Agglutination Testi (MAT) uygulandı. 98 serum örneğinin, %80.61 *toxoplasmosis*, %34.69 *listeriosis* and %6.12 *brucellosis* yönünden seropozitif olduğu tesbit edildi. Bu sonuçlar ile Van yöresi keçilerinde *Toxoplasmosis*'in, *listeria* ve *brucella*'ya göre çok daha yaygın olduğu görüldü.

Anahtar kelimeler: *Toxoplasma*, *Listeria*, *Brucella*, seroloji, seroprevalans, keçi

INTRODUCTION

In goat breeding, most important problem is infectious diseases. Main infectious diseases are toxoplasmosis, listeriosis and brucellosis which all cause abortion apart from other symptoms (1, 5, 9, 12, 17, 20, 28).

Toxoplasmosis is a zoonosis with an infectious reservoir encompassing all animals. As the definitive host, the domestic cat appears to be the major culprit in transmission to other animals. After infecting a new host, including acute, latent, and reactivated disease as well as congenital disease and chorioretinitis (20, 21). Acute acquired toxoplasmosis is most commonly asymptomatic, but it can range from mild symptomatology in the normal host to fulminant and fatal illness in the immunocompromised host. After developing an immunologic response to the initial acute infection, most normal hosts contain

the infection. The prevalence of seropositivity for *Toxoplasma* antibodies varies with geographic location, flock and country (9, 16, 20, 36).

Listeria monocytogenes is a gram positive, facultative, intracellular bacteria which is ubiquitous in nature and cause septicemia, mastitis, encephalitis and abortions in ruminants. It has emerged as one of the most important food-borne pathogens and occurs in variety of food items such as milk and dairy foods (23), seafood, raw as well as cooked egg (15) and foods of animal origin (7,27). The main route of transmission of *L. monocytogenes* is by ingestion of contaminated food and the disease is particularly common in ruminants fed on silage (39, 23). The authentic diagnosis of listeriosis has been made by isolation of the organism but it is time-consuming (24). The detection of antibodies as an indirect indication of infection among animals and man have the advantage of screening large population in a

relatively short time. The conventional tests employ crude listerial proteins such as heat-killed, trypsinized or cold-extracted antigens to detect listerial antibodies through a battery of serological tests such as the complement fixation test, agglutination, immunoprecipitation and passive immunohaemolysis (2,22,25,29).

Brucellosis is also major zoonotic problem in many countries, and its eradication in the animals is a necessary step to control the human disease. Brucellosis in goats, caused by *Brucella melitensis*, frequently results in abortions and diminished levels of milk production (10, 17, 32). Brucellosis can be considered as a great challenge to the development of dairy production in developing countries. It is one of the most economically devastating diseases, which causes great losses among the offspring and causes health problems in the rural and urban population, due to either contact with the infected materials or consumption of the contaminated dairy products (1, 10, 19, 17). Complement Fixation Test (CFT), Rose Bengal Test (RBT), and the Serum Agglutination Test (SAT) are among the most useful tests for routine diagnosis (10,17,28,30). Furthermore, currently brucella Micro Agglutination Test (MAT) has been used to detect brucellosis serologically in this country which found to be quite specific and sensitive in the serological diagnosis of brucellosis (6,10,14,17).

Goat production in this region is an important livelihood source. The region is a border city which from time to time animal crossing from other countries such as Iran, Iraq and can not be controlled properly. Thus, epidemic in this region occur more often. Problems effecting goats health is also affecting their owners economically. Therefore, in this study the seroprevalence of the most important diseases causing abortion in goats were aimed to investigate.

MATERIALS AND METHODS

Blood Samples

A total of 98 female goats at different age were examined clinically and blood samples were taken. Serum samples were obtained from these blood samples and stored at -20°C until tested. These goats were chosen because abortion in this flocks were observed in the earlier pregnancy

period. All the tests for toxoplasmosis, listeriosis and brucellosis in the serum samples taken in the present study were carried out by the Laboratories of Refik Saydam National Hygiene Center, Department of Communicable Diseases Research, Ankara.

Serological assays

All of the sera were tested for antibodies against *Toxoplasma gondii*, *Listeria monocytogenes* and *Brucella melitensis*.

Sabin-Feldman Dye Test (SFDT) for Toxoplasmosis

Serum samples were tested for anti-T.gondii antibodies in fourfold dilutions (1/16; 1/64; 1/256; 1/1024) using the standard Sabin-Feldman dye test (SFDT) as routinely performed according to the modified method of Feldman and Lamb (16). The SFDT result was regarded as positive if more than 50% of tachyzoites did not accept the dye (unstained) at $\geq 1:16$ examined under the light microscope (x 400).

Antigen preparation and agglutination test for Listeriosis

An antibody titration test to detect antibodies for *L. monocytogenes* was carried out according to the method described by Osebold et al. (29). The test antigen used in the present study was prepared in the Laboratories of Refik Saydam National Hygiene Center, Department of Communicable Diseases Research, and the assay was carried out in 3 steps. For the first step, the whole cell antigens were prepared from *Staphylococcus aureus* (ATCC 29213) strains by the Osebold method. In the second step, Listerial antigens were prepared from *L. monocytogenes* 1/2a, 1/2b, 4b, 4c and 4d strains and were combined in the same suspension. In the last step an agglutination test was performed after the absorption of sera samples with *S. aureus* antigen.

Brucella Micro Agglutination Test (MAT)

The MAT was performed as described by Baum. et al. (6). Briefly, Two-fold serial dilutions of sera, ranging from 1:2.5 to 1:40, were prepared in saline and 0.5% phenol in V-shaped microtiter plates. Fifty μl of *B. abortus* S99 antigen solution stained with Safranin-O (0.02%) composed of 5%NaCl and 0.5% phenol was added to each well containing 50 μl diluted serum and the plate was covered with a lid. The negative control wells contained phenol saline and the antigen. The results were read after 18 h of incubation at 37°C . The agglutination results were considered as negative (compact red dot) or positive (large diffuse red mat). Positive and negative controls were run for each test.

Statistical Analysis

Descriptive statistic and X^2 test were used in the serum samples to determine positivity and mix antibody titers (11).

RESULTS

Clinical findings

Clinically examined goats had no specific clinical signs of the diseases. But in their history; the flocks suffered from abortus in their previous pregnancy period.

Serological findings

According to Sabin-Feldman Dye Test (SFDT); out of for 98 goat 79 (80.61%) were seropositive for toxoplasmosis at $\geq 1:16$ dilutions. Seropositivity distribution at different dilutions were 42 (42.85%) at 1/16 dilution, 27 (27.55%) at 1/64 dilution, 7 (7.17%) at 1/256 dilution and 3 (3.06%) at 1/1024 dilutions. Furthermore, 34 samples (34.69%) were seropositive for listeriosis according to Osebold Agglutination Test (OAT). MAT for brucellosis also revealed 6 (6.12%) seropositivity at $\geq 1/20$ dilutions. Additionally, 32(32.6%) seropositivity to both toxoplasmosis and listeriosis, 7(7.14%) seropositivity to both toxoplasmosis and brucellosis, 3 (3.06%) seropositivity to both listeriosis and brucellosis, 3 (3.06%) seropositivity to all toxoplasmosis, listeriosis and brucellosis were determined.

DISCUSSION

Infectious diseases causing abortus such as toxoplasmosis, listeriosis and brucellosis are important diseases for goats (1,10,20). They cause important economical losses, not only by abortus but also reduction in milk production. Especially the goat production is one of the most important income for the farmers. Their importance does not come only from their effect on animal, they also contaminate human beings (21,33,34).

Although seropositivity to an organism does not translate into verification that the animal was clinically affected by that organism, the infectious diseases; toxoplasmosis, listeriosis and brucellosis seem to be widespread among goat flock from the region of Van and probably represents an important factor that contributes to the decreased productivity of those animals (35,36). Most probably decreased goat production in Turkey has been occurring year by year due to such diseases. Unfortunately, there aren't many

studies concerning these diseases surveillance. Therefore, these disease prevalence need to be put forward to combat and to make proper eradication programs.

Studies concerning toxoplasmosis have been made on several animal species including on goats world wide (21,26,36). Several serological tests have been used to detect antibodies against toxoplasmosis. Many tests are available for the detection of specific antibodies to toxoplasmosis (16, 20, 26). One of them is SFDT, which is used for diagnostic purpose and considered a gold standard test. In the present study, seropositivity to toxoplasmosis was found to be 80.61% using this test. The frequency of toxoplasmosis infection is extremely variable in the different regions of the world. Several studies have been performed on toxoplasmosis in different parts of the world and prevalence of seropositivity was found 11.6-96% in goats (9, 20, 26). In the studies on the seroprevalence of toxoplasmosis in the different regions of Turkey had been reported between 12.1-63.1% in goats (35, 36). It can be seen from our results that findings reported in this study with regard to toxoplasmosis was quite high in comparison to results given for average Turkey's results. This could be due to the animals owners unawareness to the vaccination programme and also the regions characteristics. The region is border to Iran and Iraq which illegal animal trade occur from time to time. Therefore, high seropositivity could also be relied on this situation.

In the present study seropositivity to listeriosis was found to be 34.69% in goats. Epidemiological studies have revealed that only *Listeria monocytogenes* and only strains belonging to serotypes 1/2a, 1/2b and 4b were implicated in 90% of outbreaks of listeriosis (18). Therefore in the present study antibodies against *Listeria monocytogenes* were investigated using OAT test. In recent years PCR and ELISA tests have widely been carried out in the diagnosis of listeriosis (3,5,12,13). But, OAT test has also been used safely especially in epidemiological screenings (29). Listeriosis is mainly transmitted by ingestion of contaminated food and the disease is particularly common in ruminants fed on silage (8,31). However, goats used in the present study have never been had silage according to anamnesis. According to different serological test; the seroprevalence of listeriosis in goats have been reported to be between 14.52% and 41.13% (4, 5, 31). Findings reported in

the present study were also in these limits. In a study carried out by Tütüncü et al. (37) found the seroprevalance of listeriosis in cattle as 28.5% in the region of Van. In addition, Erdoğan et al.(13), studied seroprevalance of listeriosis in cattle and found 88.7% using ELISA in the region of Kars. But, we couldn't sight any seroprevalance studies on goat in Turkey. Therefore, findings in this study should be considered as novel results. Furthermore, studies concerning listeriosis in this region (which is a border region) have not been investigates before.

Brucella melitensis is the most common agent of caprine and human brucellosis. Therefore, small ruminants are considered the main source of human infection (26,32,34). Brucellosis is worldwide disease particularly in Near East countries, middle East, Iran, Iraq, Turkey including province of Van (32, 34). It is one of the most economically devastating disease, which causes great losses among the offspring and causes health problems in rural and urban population, due to either direct contact with infected materials or consumption of the contaminated dairy products such as milk (32). Eradication of this disease in the animals is a necessary step to control the human disease. In the European Union, this disease is limited to

Mediterranean countries. Although the disease seroprevalance in sheep have been reported to be quite high in Turkey, Its serological presence in goat has been reported to be low in comparison to sheep (28,32,34). In the present study the seroprevalance of brucellosis in goats were found to be 6.12%. The test used in the present study to determine seroprevalance of brucellosis was MAT which has widely been used in Turkey for screening brucellosis (14). On the other hand, screening brucellosis in Turkey has been concentrated mainly on sheep and cattle (19). Furthermore, brucella screening on goats in our region (which is more vulnerable) have not been carried out according to our knowledge so far. Therefore, this study concerning brucellosis in this region should be considered as novel findings.

As a result, with these findings, seroprevalance of the diseases causing abortions in goats were found to be quite high in this region. Furthermore, results showed that the seroprevalance of toxoplasmosis was the highest. These diseases are very important with regard to economical losses. Therefore, to eradicate such disease and to reduce economical losses, more detailed studies should be made in this region to make proper and effective challenge.

References

1. Al-Majali, A.M. (2005): Seroepidemiology of caprine Brucellosis in Jordan. *Small Ruminant Research*. 58; 13-18.
2. Aslan, V., Turgut, K., Kaya, O., Sevinç, M. (1991): Sığırlarda Listeriosis Olgusu. *Hayvancılık Araştırma Derg.* 1 (1): 37-39
3. Baetz, A. L., Wesley, I.V. (1995): Detection of anti-listeriolysin O in dairy cattle experimentally infected with *Listeria monocytogenes*. *J Vet Diagn Invest*. 7(1):82-6.
4. Barbuddhe, S.B., Chaudhari, S.P, Malik, S.V.S. (2002): The occurrence of pathogenic *Listeria monocytogenes* and antibodies against listeriolysin-O in buffaloes. *J Vet Med B Infect Dis Vet Public Health*. 49 (4): 181-4.
5. Barbuddhe, S.B., Malik, S.V.S., Bhilegaonkar, K.N., Gupta, L. K. (2000): Isolation of *Listeria monocytogenes* and anti-listeriolysin O detection in sheep and goats. *Small Ruminant Research*. 38; 151-154.
6. Baum, M., Zamir, O., Bergman-Rios, R., Katz, E., Beider, Z., Cohen, A., Banal, M. (1995): Comparative Evaluation of Microagglutination Test and Serum Agglutination Test as Supplementary Diagnostic Methods for Brucellosis. *J. Clin Microbiol*. 33: 2166-2170
7. Berche, P., K. A. Reich, M. Bonnicon, J. L. Beretti, C. Geoffroy, J. Raveneau, R. Cossart, J. L. Gaillard, P. Geslin, H. Kreis, and M. Veron, 1990: Detection of anti-listeriolysin O for serodiagnosis of human listeriosis. *Lancet*. 335, 624-627.
8. Boerlin, P., Boerlin-Petzold, F., Jemmi, T. (2003): Use of Listeriolysin O and Internalin A in a Seroepidemiological study of Listeriosis in Swiss Dairy Cows. *J of Clinical Micro*. 1055-1061.
9. Borde, G., Lowhar, G and Adesiyun A.A.(2006): *Toxoplasma gondii* and *Chlamydia abortus* in caprine Abortions in Tobago: a sero-Epidemiological Study. *J. Vet. Med. B*. 53, 188-193.
10. A. M., and Commander, N. J. (2005): A review: Brucellosis- new aspects of an old disease. *J. of Applied Microbiology*. 98; 1270-1281.
11. Dean, A.G., Dean, J.G., Coulombier, D., Berendel, K.A., Smith, D.C., Burton, A.H, Dicker, R.C., Sullivan, K.M., Fagan, R.F., Arner, T.G. (1994): *Epi-Info Version 6: A Word Processing Database and Statistics Program for Epidemiology on Microcomputers*. Center for Disease Control and Prevention. Atlanta. Georgia. USA.

12. Donachie, W., Low, J.C. (1995): Ovine Listeriosis. The Veterinary Annual. 35, 304-312.
13. Erdoğan, H.M, Gökçe, G., Gökçe, H.İ, Kırmızıgül, A.H, Güneş, V., Sural, E., Yılmaz, K. (1999): Kars yöresindeki sığırlarda *Listeria monocytogenes* enfeksiyonlarının ELISA yöntemi ile araştırılması. Kafkas Ü. Vet. Fak. Derg. 5(1): 43-46.
14. Esendal, Ö.M., Yardımcı, H., Keskin, O., Altay, G. (2001): Sığır, Koyun ve Keçi Brucellosis'inin Serolojik Tanısında Konvansiyonel Testler ve Coombs Testinin Kullanılması. Ankara Üniversitesi Veteriner Fakültesi Dergisi. 48: 97-102.
15. Farber, J. M., and Peterkin, P. I (1991): *Listeria monocytogenes*, a food-borne pathogen. Microbiol. Rev. 55, 476-511.
16. Feldman, H.A and Lamb, G. A. (1966): A micromodification of the Toxoplasma Dye Test. J Parasitol; 52: 415.
17. Garin-Bastuji, B., Blasco, J. M. Marin, C., Albert, D. (2006): The diagnosis of brucellosis in sheep and goats, old and new tools. Small Ruminant Research. 62; 63-70.
18. Gasanov, U., Hughes, D., and Hansbro, M. P: (2005): Methods for the isolation and identification of *Listeria* spp. and *Listeria monocytogenes*: a review. FEMS microbiology Reviews. 29; 851-875.
19. Gürtürk, K., Alan, M., Boynuvara, B., Solmaz, H. (1994): Van ve Yöresinde Koyun ve Sığır Brucellozisinin İnsidensi Üzerinde Sero-Epidemiyolojik Araştırmalar. Yüztüncü Yıl Üniversitesi Veteriner Fakültesi Dergisi; 5: 121-125.
20. Hill, D., Dubey, J. P. (2002): *Toxoplasma gondii*: transmission, diagnosis and prevention. Clin Microbiol Infect. 8: 634-640.
21. Jittapalpong, S., Sangvaranond, A., Pinyopanuwat, N., Chimnoi, W., Khachaeram, W., Koizumi, S., Maruyama, S. (2005): Seroprevalence of *Toxoplasma gondii* infection in domestic goats in satun province, Thailand. Veterinary Parasitology. 127; 17-22.
22. Lhopital, S., J. Marty, P. Pardon, and P. Berche, (1993): Kinetics of antibody production against listeriolysin-O in sheep with listeriosis. J. Clin. Microbiol. 31, 1537-1540.
23. Low, J.C., Donachie, W. (1997): Reweiv of *Listeria monocytogenes* and Listeriosis. The Veterinary Journal, 153 (1): 9-29.
24. Low, J. C., and Donachie, W. (1991): Clinical and serum antibody responses of lambs to infection by *Listeria monocytogenes*. Res. Vet. Sci. 51, 189-192.
25. Low, J. C., R. C. Davies, and W. Donachie, (1992): Purification of listeriolysin-O and development of an immunoassay for diagnosis of listeric infections in sheep. J. Clin. Microbiol. 30, 2705-2708.
26. Masala, G., Porcu, R., Madau, L., Tanda, A., Ibba, B., Satta, G., Tola, S. (2003): Survey of ovine and caprine toxoplasmosis by IFAT and PCR assays in Sardinia, Italy. Veterinary Parasitology. 117; 15-21.
27. McLauchlin, J. (1996): The Relationship between *Listeria* and Listeriosis. Food Control. 7(4-5): 187-193.
28. Minas, A. (2006): Control and eradication of brucellosis in small ruminants. Small Ruminant Research. 62; 101-107.
29. Osebold, J., Aalund, O., Chrisp, C. E. (1965): Chemical and immunological composition of surface structures of *Listeria monocytogenes*. J. Bacteriol. 89: 84-86.
30. Portanti, O., Tittarelli, M., Febo, D., Luciani, M., Mercante, M. T., Conte, A., and Lelli, R. (2006): Development and Validation of a competitive ELISA Kit for the serological diagnosis of ovine, caprine and Bovine Brucellosis. J. Vet.Med. B. 53; 494-498.
31. Rekha Bhanu, V., Malik, S. V. S., Chaudhari, S. P., Barbuddhe, S. B. (2006): Listeriolysin O-based diagnosis of *Listeria monocytogenes* infection in experimentally and naturally infected goats. Small Ruminant Research. 66; 70-75.
32. Shareef, J.M. (2006): A Review of Serological Investigations of Brucellosis among Farm Animals and Humans in Northern Provinces of Iraq (1974-2004). J. Vet.Med. B. 53; 38-40.
33. Skovgaard, N., and C. Morgen, (1988): Detection of *Listeria* spp. in faeces from animals, in feeds, and in raw foods of animal origin. Int. J. Food Microbiol. 6 ,229-242.
34. Taleski, V., Zerva, L., Kantardjiev, T., Cvetnic, Z., et.al. (2002): An overview of the epidemiology and epizootology of brucellosis in selacted countries of central and southeast Europe. Veterinary Parasitology. 90; 147-155.
35. Tütüncü, M., Akkan, H. A., Babür, C., Ayaz, E., Karaca, M. (2001): The seroprevalance of *toxoplasma gondii* in sheep detected by Sabin Feldman Dye Test in the region of Van, Turkey. Y.Y.U. Vet. Fak. Derg. 12 (1-2): 33-35.
36. Tütüncü, M., Ayaz, E., Yaman, M., Akkan, H. A. (2003): The seroprevalance of *toxoplasma gondii* in sheep, goats and cattle detected by indirect hemagglutination (IHA) test in the region of Van, Turkey. Indian Vet. J 80: 401-403.
37. Tütüncü, M., Solmaz, H., Akkan, H.A., Karaca, M., Ağaoğlu, Z. (2005): The Investigation of *Listeria monocytogenes* in cattle detected by ELISA test in the region of Van, Turkey, Indian Vet. J, 82, 926-928.
38. Ueno, H., Yokota, K., Arai, T., Muramatsu, Y., Taniyama, H., Iida, T., Morita, C. (1996): The prevalence of *Listeria monocytogenes* in the environment of dairy farms. Microbiol Immunol. 40(2):121-24.
39. Unnerstad H, Romell A, Ericsson H, Danielsson-Tham ML, Tham W. (2000): *Listeria monocytogenes* in faeces from clinically healthy dairy cows in Sweden. Acta Vet Scand. 41(2):167-71.